ROSBRED

Enabling marker-assisted breeding in Rosaceae

Phenotyping Protocol for Sweet Cherry (Prunus avium L.) to Enable an **Understanding of Trait Inheritance**

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Introduction

The USDA Specialty Crop Research Initiative-funded RosBREED project has the objective of enabling marker-assisted breeding (MAB) in the economically important agricultural family of Rosaceae. To standardize and increase the accuracy of MAB in Rosaceae, it is necessary to characterize many horticultural and fruit quality traits in representative germplasm. A well-developed comprehensive phenotyping protocol for productivity, fruit quality and horticulturally objectionable traits, developed for use in the Washington State University (WSU) Sweet Cherry Breeding Program, is described and correlations among selected traits presented.

Materials and Methods

This study was carried out at Washington Sate University, Irrigated Agriculture Research and Extension Center, Prosser. Trees were maintained at the Roza farm under standard orchard conditions. Data collected (see Table 1 for protocols) from a sweet cherry 'Crop Reference Set' and 'Breeding Pedigree Set' between 2010 and 2012 were subjected to ANOVA and phenotypic correlations among selected traits were also calculated.

Results

A strong positive correlation (Figure 1) was observed between harvest date and fruit weight and fruit firmness (+0.26, p<0.0001 and +0.39, p<0.0001, respectively), with later-maturing varieties tending towards larger and firmer fruit. Also, fruit weight has a positive and significant association with pedicel-fruit retention force with a value of 0.43 (p<0.0001). On the other hand, soluble solids content showed a negative relationship with fruit weight and firmness (-0.34 and -0.20, respectively). The box plot for representative traits is presented in Figure 2. There were significant (p<0.05) yearly differences in pedicel fruit retention force, soluble solids content, bloom time and harvest time while titratable acidity was fairly consistent across years.

Figure	• 1: Phe	notypi	ic corr	elatior	ns amo	ng 20 g	sweet o	cherry	traits	at WSI	J Pros	ser IAF	REC. W	/A base	ed on 3	vears	data c	ombin	ed ^{1,2,3}
SSC	TA	FL	FW2	FWt	PL	PW2	PWt	FS	CMS	CBS	CFI	ShF	Fir	FirB	HD	HT	BD	BT	
ns	ns	0.38	<u>0.41</u>	0.43	0.24	<u>0.19</u>	0.27	ns	ns	ns	ns	ns	0.31	0.20	0.16	0.16	ns	ns	PFRF
	<u>0.33</u>	<u>-0.41</u>	-0.38	-0.34	-0.25	-0.15	-0.15	-0.18	<u>0.29</u>	ns	ns	ns	-0.20	-0.13	-0.18	-0.17	ns	ns	SSC
		-0.14	ns	ns	-0.17	ns	ns	ns	ns	ns	ns	ns	ns	ns	<u>0.23</u>	<u>0.23</u>	ns	ns	ТА
			<u>0.90</u>	<u>0.89</u>	<u>0.71</u>	<u>0.38</u>	<u>0.49</u>	<u>0.24</u>	-0.15	ns	0.14	<u>0.25</u>	<u>0.41</u>	<u>0.30</u>	<u>0.24</u>	<u>0.23</u>	ns	ns	FL
				<u>0.95</u>	<u>0.57</u>	<u>0.45</u>	<u>0.50</u>	<u>0.23</u>	ns	-0.29	ns	0.18	<u>0.43</u>	<u>0.31</u>	<u>0.24</u>	<u>0.23</u>	ns	ns	FW2
					<u>0.61</u>	<u>0.44</u>	<u>0.53</u>	0.15	ns	-0.31	ns	0.19	<u>0.39</u>	<u>0.21</u>	<u>0.26</u>	<u>0.25</u>	ns	ns	FWt
						<u>0.29</u>	<u>0.60</u>	0.13	ns	ns	0.14	<u>0.21</u>	<u>0.28</u>	<u>0.20</u>	<u>0.20</u>	<u>0.20</u>	ns	ns	PL
							<u>0.43</u>	ns	ns	ns	ns	ns	0.18	ns	0.16	0.15	ns	ns	PW2
								ns	ns	ns	ns	ns	<u>0.27</u>	0.18	<u>0.26</u>	<u>0.26</u>	ns	ns	PWt
									ns	ns	0.15	ns	ns	ns	ns	ns	ns	ns	FS
										•	<u>0.66</u>	ns	-0.16	ns	-0.19	0.19	ns	ns	CMS
¹ Correlation values significantly different at $p < 0.0001$ level are underlined								ns	ns	ns	ns	ns	ns	ns	ns	CBS			
p < 0.001 level in italics, and all remaining values are at $p < 0.01$ level.										ns	ns	ns	<u>-0.26</u>	<u>0.26</u>	ns	ns	CFI		
Values not significant at any of these levels are identified as non-significant (ns).0.190.21nsns									ns	ShF									
² PFRF=pedicel-fruit retention force, SSC=soluble solids content, TA= titratable acidity, FL=fruit length,									Fir										
FW2=fruit width, FWt=fruit weight, PL=pit length, PW2=pit width, PWt=pit weight, FS=freestone, CMS=mahogany skin color, CBS=blush skin color, CFl=flesh color, ShF=fruit shape, Fir=Firmness, FirB=bulked fruit firmness, HD=hervest date, HT=hervest time, BD=bloom date, BT=bloom time,											FirB								
F11B=DU	$\underline{0.99} \underline{0.26} \underline{0.24}$										HD								
³ Correlations were calculated with the corr procedure in SAS version 9.2 (SAS Institute Inc., Cary, NC).												<u>0.24</u>	<u>0.23</u>	HT					
																		<u>0.95</u>	BD



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Correlations revealed through statistical analysis of data collected in multiple harvest seasons can be used to elucidate trends in trait associations. Measuring either bulked fruit firmness or firmness based on five fruit only, as well as, either bloom/harvest time in julian days or growing degree days, will streamline the phenotyping protocol and improve cost effectiveness. The significant and positive correlations between SSC and TA as well as between harvest date, fruit size and firmness may suggest a genetic linkage of genes controlling these traits. Use of this standardized phenotyping protocol will enable comparisons of data between years and programs and aid in identifying sweet cherry regions that control trait variation.



CONCLUSIONS

WASHINGTON STATE

Table 1: Phenotypic traits, trait code, unit of measure and description used to categorize sweet cherry accessions

Trait code Unit

of measure	Description
Productiv	ity traits
day	The number of days from January 1 to date of 50% bloom
ng Degree	GDD accumulated from January 1 to date of 50% bloom calculated with a base temperature of 4.4 °C
day	The number of days from January 1 to harvest
ng Degree	GDD accumulated from January 1 to harvest calculated with a base temperature of 4.4 °C
Fruit qual	ity traits
1	Measured by compression (cheek to cheek) on a Firmtech (BioWorks, Kansas USA) using 5 TE ¹
	Measured by compression (cheek to cheek) on a Firmtech (BioWorks, Kansas USA) using 20 BF ²
	This is the force required to pull the pedicel off the fruit measured with a pull force gauge (Imada DS-2). 5 TF samples are used.
	Juice from 5 TF samples pulled together is used and Brix% is recorded on a refractometer (Atago PAL-1)
	Juice from BF samples is used
	Measured with a 10 ml strained juice from BF on an autotitrator (Metrohm, Florida, USA) titrated to pH endpoint 8.2
	Same as above
	Average weight of 5 TF
	Average weight of 20 BF t
	Average of 5 IF
	Measured from stem to blossom scar, average of 5 IF
	TF
	Measured from cheek to cheek, average of 5 TF
	Using the CTFILcolor chart and recording 1 to 7
	1=0-25%, 2=26-50%, 3=51-75% and 4=76-100%
	Ratio of Fruit Length and Width1; 1=kidney, 2=flat-round, 3=round, 4=elongate, 5=cordate (heart-shaped)
	Visual Flesh Color Card rating 1-5, $1 = \text{cream}$, $2 = \text{pink}$, $3 = \text{red}$, $4 = \text{red}$ purple and $5 = \text{mahogany}$
	Pit cling: 1= no cling, 2=slight cling, 3=semi cling, 4=cling, 5=total cling, average of 5 TF
	Average pit weight of 5 TF
	Pit length from point to point, average of 5 TF
	Pit width perpendicular to width 1, average of 5 TF
ruit objectio	
ruit objectio	Calculated as the percentage of fruit with pitting out of a
	Calculated as the percentage of fruit with splits out of a total of 20 fruit sampled
j	 0=no visible symptoms, 1=very slight infection on leaves, 2=slight infection on leaves, occasionally on shoots, 3=moderate infection on leaves and/or shoots, 4=heavy infection on leaves and/or shoots and 5=very high infection on leaves, most shoots damaged
1	0=no necrosis, 1=line of infection, $2 = \le 50\%$ of the leaf, $3 = >50\%$ of the leaf and $4 = total$ necrosis